

Research article

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An Eco-friendly Approach - Synthesis and Characterization of CuO nanoparticles

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ABSTRACT

Nanoparticles hold excellent and attractive properties due to their small sizes, large surface area, free hanging bonds and superior reactivity. Nanoparticles (NPs) are a part of nanomaterial that is defined as a single particle 1-100 nm in diameter. From last few years, nanoparticles have been a conventional material for the development of new cutting-edge applications in communications, energy storage, sensing, data storage, optics, transmission, environmental protection, cosmetics, biology, and medicine due to their essential optical, electrical, and magnetic properties. Synthesis of nanoparticles can be performed using a number of routinely used chemical and physical methods. Chemical synthesis methods for NPs include emulsion solvent extraction method, double emulsion, and an evaporation method, salting out method, emulsion diffusion method, and solvent displacement/precipitation method. But, industrial-scale production of NPs has familiarized a new kind of pollution into the environment. Thus, there is a need for "green synthesis" that includes a clean, safe, eco-friendly and environmentally nontoxic method of nanoparticle synthesis. Various reports emphasize that plant-based nanoparticles had the valuable impact on agriculture, pharmaceuticals, drug delivery and production of other commercial goods. Use of plant extracts as natural reducing, capping and stabilizing agents have been attained considerable progress. Green synthesis method avoids inert gases, high pressure, laser radiation, high temperature, toxic chemicals etc. as compared to the conventional method like sol-gel technique method, laser ablation method, inert gas condensation method, chemical reduction method etc. The structures, morphology, optical properties, surface area and thermal behavior of these fabricated nanoparticles were characterized by X-ray Diffraction (XRD), Scanning Electron Microscopy (SEM), Ultraviolet-visible spectroscopy (UV-vis), Photoluminescence (PL)/fluorescence spectroscopy and Fourier transform infrared spectroscopy (FTIR) analysis. This review is a comprehensive study of the synthesis, structure and physiochemical characterization methods used for synthesis of CuO NPs via green approach.

KEY WORDS: Green synthesis of nanoparticles, FT-IR, XRD, SEM, CuO NPs.

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INTRODUCTION

"Nanotechnology is the application of science to control matter at the molecular level"¹.Nanotechnology deals with the science and engineering of creating functional materials and devices between 0.1 to 100 nanometer scale in any dimension and is the most commercially viable technology of the 21st century. Tremendous growth in nanotechnology has opened up novel fundamental and applied frontiers in materials science and engineering, such as Nano biotechnology², bio nanotechnology³, quantum dots⁴, surface-enhanced Raman scattering (SERS)⁵, and applied microbiology. Nanoparticles have been intensively studied over the last decade due to their characteristics: physical, chemical, electronic, electrical, mechanical, magnetic, thermal, dielectric, optical and biological properties^{6,7}. Non-Toxic property of nanoparticles has been utilized for cancer cell treatment.⁸⁻¹⁰. Metal oxide nanoparticles are of much interest because of their unique optical, electronic and magnetic properties. The transition metal oxide are acting as semiconductors and can be utilized as magnetic storage media, solar energy transformation, electronics, gas sensors and catalyst ¹¹⁻¹⁵. Numerous physical and chemical methods have been broadly used to produce Nano crystalline copper oxide such as micro emulsion method¹⁶, arc-submerged nanoparticle synthesis system¹⁷, flame-based aerosol methods¹⁸, nonchemical¹⁹, hydrothermal²⁰ and solid-state techniques²¹. When hazardous chemicals are used on the surface of nanoparticles and non-polar solvents in the synthesis procedure it decreases the chances of use of Nano materials in clinical field. Hence ecofriendly methods for synthesis are raise as it gives biocompatible and non-toxic nanoparticles. The most prominent eco-friendly method of recent interest is green approach. These approaches focus on utilization of environment-friendly, cost-effective and biocompatible reducing agents for synthesis of nanoparticles. Numerous biomaterials used for synthesis of NPs, but plant are very less toxic amongst them and nanoparticles produced by plants are more stable, are of various sizes and shapes²². The use of harsh chemicals such as hydrazine hydrate, sodium borohydride, dimethylformamide, ethylene glycol, and so on producing very toxic effects. To overcome this problem, biosynthetic and environment friendly methods are employed for synthesis of NPs. Many of these reducing agents have been associated with environmental toxicity or biological hazards. The synthesis of metal nanoparticles using inactivated plant tissue, plant extracts, exudates, and other parts of living plants is a modern alternative for their production²³Cu, Cu₂O and CuO NPs.Copper nanoparticles caneasily oxidize to form copper oxide. If the application requires the copper nanoparticles to be protected from oxidation, the copper NPs are usually encapsulated in organic or inorganic material such as carbon and silica ²⁴⁻²⁷. Review of literature revealed that synthesis of CuONPs using microorganisms and plant extract has been unexplored; there are only a very few reports on the use of yeast, fungi, bacteria or plant extract for synthesizing CONPs²⁸⁻³⁰. Different analytical techniques were used including UV–visible spectroscopy (UV–vis), Fourier transform infrared spectroscopy (FTIR), transmission electron microscopy (TEM) and X-ray diffraction analysis (XRD) for characterization of CONPs. Furthermore, the bacterial effect of CONPs was also analyzed by disc diffusion method. The current review highlights the information about different methodology and plant extract used for synthesis of copper oxide nanoparticles and further this information can be employed for understanding the properties and behavior of CuO nanoparticles in different applications.

MATERIALS AND METHODS

1. Green synthesis of copper oxide nanoparticles using caricapapaya.

Sankar et al., worked on synthesis of Copper oxide nanoparticles using cupric sulphate. Phytochemical present in the plant leaves act as reducing agent and able to reduce copper ions. Nanoparticles has been prepared via green approach. Further the confirmation of copper oxide nanoparticles has been done through XRD and size and shape confirmation via SEM. Synthesized nanoparticles were rod shape and size distribution was between 100-150nm. Further the application of nanoparticles in the field of dye degradation was observed.³¹.

2. Synthesis Copper Oxide Nanoparticles using Malvasylvestris Leaf Extract.

Awwad*et al.*, used leaves of *Malvasylvestris* were collected in and around the campus of Royal Scientific Society, El Hassan Science City, Amman, Jordan. They synthesized copper oxide nanoparticles from *Malvasylvestris* and plant extract was treated with copper chloride dehydrate. Further XRD and SEM results clarify the confirmation of copper oxide nanoparticles. CuO NPs were further used as antimicrobial agent ³².

3. Green synthesis of copper oxide nanoparticles using Gum karaya.

M.Cernik*et al.*, reported the antimicrobial effect of copper oxide nanoparticles using *Gum karaya*. Plant leaves act as reducing agent and phytochemicals present plant reduce copper hydroxide to nano crystalline copper oxide nanoparticles. Antimicrobial activity checked with different gram positive and negative bacterial strains and study shows that nanoparticles have good antimicrobial property³³.

4. Green synthesis of copper oxide nanoparticles using Tabernaemontana divaricate leaf extract.

M. Nasrollahzadeh reported the synthesis of palladium copper oxide nanoparticles from leaves of *Tabernaemontana*. Polyphenols present in the plant can reduce the copper sulphate to copper oxide nanoparticles. The synthesized nanoparticles were further used as catalyst to 4-Nitrophenol³⁴.

5. Green synthesis of CuOusing brown alga extract Bifurcaria bifurcate.

Y. Abbound*et al.*, reported the antimicrobial activity of copper oxide nanoparticles synthesized from brown algae. Phytochemical of brown algae reduce Copper (II) sulphate to copper oxide nanoparticles.Nanoparticles of size range 5 nm to 50nm shows effective antimicrobial activity.³⁵.

STRUCTURE AND PHYSICO-CHEMICAL CHARACTERIZATION

Sr.	Author	Year of	Type Of	Type Of	Observation	Ref.
No.	Name	Publication	Nanoparticle	Characterization		no.
1.	Amrut. S.	2010	CuO NP	SEM	Rectangular	36
	Lanje et. al			TEM	Size- 5-6 nm	
				XRD	Crystalline size 8 nm	
2.	Υ.	2013	CuO NP	TEM	Size- 5-45 nm. Average	35
	Abbounand		using		size-20.66. Spherical and	
	et. al		Bifurcariybifu		small percentage of	
			rcate		elongated shaped particle.	
				FT-IR	Peaks at 3413, 1730, 1625,	
					1103, 1033, 3000 and 1400	
					cm ⁻¹ .	
				XRD	20 values 29.4°, 36.8°,	
					42.1°, 61.9°, and 77.6°.	
3.	ThekkaePadil	2013	CuO NP	SEM	Small needle like structure	33
	and Cernic		using Gum	TEM	Size- 2-10 nm	
			karaya	FT-IR	Peaks at 3150, 3025, 1740,	
					1600, 1400, 1250, 1030,	
					525, 580, 675 cm ⁻¹ .	
				XRD	20 values 32.47°, 35.49°,	
					38.68°, 48.65°, 53.36°,	
					58.25°, and 61.45°.	
4.	RenuSanker	2014	CuO NP	FT-IR	Peaks at 3444.4, 2926.79,	31
	et. al		using Carika		2858.10, 2363.93, 1880.01,	
			рарауа		1636.69, 1391.64, 1087.31,	
					780.57, 691.12, 473.25	
					cm ⁻¹	
				XRD	20 values 32.816°,	
					38.842°, 61.403° and	

Table No. 1: "Characterization of Nanoparticles."

					71.189°.	
5.	Awwad,	2015	CuO NP	SEM	Spherical, size- 5-30 nm.	32
	A.M. et. al		using	FT-IR	Peaks at 3444, 3398, 2924,	
			Malvasylvestr		2846, 1734, 1604, 513 cm	
			is		1	
				XRD	2θ values of 32.49°, 35.49°,	
					38.96°, 48.73°, 53.45°,	
					58.34°, 61.53°, 65.79°,	
					66.25°, 72.43°, and 75.03°.	
					Crystalline size 14 nm.	
6.	H. Raja	2015	CuO NP	SEM	Spherical	37
	Naika et. al		using	TEM	Spherical, size- 5-10 nm	
			Gloriosasupe	XRD	Crystalline size 8 nm	
			rba		5	
7.	Lily Riya and	2015	CuO NP	FT-IR	Peaks at 622 cm ⁻¹	38
	Mary George		using	XRD	20 values 29.50°, 36.38°,	1
			Camellia		42.29°, 52.47°, 61.43°,	
			sinensis		73.62° and 77.50°.	
8.	MarymBordb ar <i>et. al</i>	2016	CuO NP using Rheum palmatum	SEM	Spherical, size- 30nm.	39
0.				TEM	Spherical, size-10-20 nm.	
				FT-IR	Peaks at $400-600 \text{ cm}^{-1}$.	
			1	XRD	20 values 35.39°, 38.77°,	
				me	48.85° and 61.69°.	
9.	ILL-MIN	2016	CuO NP using Ecliptaprostr ata	SEM	Spherical, hexagonal and	40
9.	Chung et. al			5 Divi	cubical. Size- 28-105 nm.	
				TEM	Spherical, size- 28-45 nm.	
				FT-IR	Peaks at 3333, 2917, 1615,	1
				1 1-11	1048 cm^{-1} .	
				XRD	20 values 31.65°, 45.42°,	
				Mill	66.14° and 75.14°.	
					00.11 and 75.11.	
10.	Long-Bao Shi	2017	CuO NP	SEM	Spherical shape	41
	et. al		using Cassia	TEM	Spherical, size- 23.70 nm	
			auriculata	FT-IR	Peaks at 3385, 2922, 1720,	
					1612, 1033, 668 and 601	
					cm ⁻¹ .	
				XRD	20 values 32.54°, 35.56°,	
					38.77°, 48.74°, 53.53°,	
					58.37°, 61.56°, 66.29° and	
					68.17°.	
1.1		2017				40
11.	Mangesh S.	2017	CuO NP	SEM	Spherical, size- 18-20 nm.	42
	Jadhav et. al		using Malus	TEM	Size- 18-20 nm.	
			Domestica	FT-IR	Peak values 3448.20,	
					2966.80, 2924.59, 2854.40,	
					1751.10, 1630.97, 1465.58,	
					1402.41, 1265.41, 1028.17	
					and 711.18 cm ⁻¹ .	4
				XRD	20 values 23.501° and	
					42.695°.	

Literature clearly shows the characterization of CuO NPs by using different methods. Scanning Electron Microscopy (SEM) and Transmission Electron Microscopy (TEM) confirms the size and shape of nanoparticles. Size of nanoparticles between 1-100nm is considered for excellent catalytic activity. Fourier-transform infrared spectroscopy (FT-IR) is utilized for the structural confirmation of molecules present inside the prepared sample. FT-IR gives the information about bond stretching in particular sample. CuO NPs shows bond stretching between 2800 -4000 cm⁻¹. X-ray Diffraction (XRD) method is used to know the crystalline structure of CuO nanoparticles. Metallic nature of CuO nanoparticles gives the $2\Theta = 42$ ° to 48 °. Current review highlights the detailed characterization and synthetic methods for CuO nanoparticles which can be helpful to understand and predict the properties of CuO NPs.

CONCLUSION

The review highlights the green chemistry approach for the synthesis of CuO nanoparticles is simple, cost effective and eco-friendly. The resultant NPs are highly stable and reproducible. Simple, eco- friendly green synthesis of CuO NPs has been reported using different leaf extract as capping and reducing agent. Here, green extract was used for the reduction of copper material at Nano scale. The present study establishes that rod or spherical shape NPs of CuO can be obtained by using these method. The characterization of CuO NPs is done by using various analytic instrument like SEM, TEM, FT-IR and XRD, which are helpful for further analysis these prepared CuO NPs has used for various applications for better human life such as it can be implemented in anti-cancer, antimicrobial, anti-oxidant activity etc.

REFERENCES

- 1. Senapati S., Biosynthesis and immobilization of nanoparticles and their applications, PhD thesis, University of Pune, 2005.
- 2. Klefenz H., Nanobiotechnology: From molecules to systems. Eng. Life Sci., 2004; 4: 211-8.
- 3. Goodsell D., Bionanotechnology: Lessons from Nature, Willey-Less, NewJersey, USA, 2004.
- 4. Chan W. and S. Nie, Colloid and Colloid Assemblies WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim: Republic of Germany: 1998.
- Z. Tian and B. Ren, Annu. Surface Enhanced Raman Spectroscopy: Imperial College: London; 2005.
- Daniel MC, Astruc D Gold nanoparticles: assembly, supramolecular chemistry, quantum-size related properties, and applications towards biology, catalysis and nanotechnology. Chem Rev, 2004; 104:293–346.
- Schmid G, Large clusters and colloids metals in the embryonic state. Chem Rev, 1992; 92:1709– 1727.

- 8. Perelshtein I, Applerot G, Perkas N, et al. CuO-cotton nanocomposite:formation, morphology, and antibacterial activity. Surface and Coatings Technology. 2009; 204(1–2):54–57.
- 9. Lee HJ, Lee G, Jang NR, Yun JH, Song JY, Kim BS. Biological synthesis of copper nanoparticles using plant extract. Nanotechnology, 2011; 1(1):371–374.
- Valodkar M, Jadeja RN, Thounaojam MC, Devkar RV, Thakore S. Biocompatible synthesis of peptide capped copper nanoparticles and their biological effect on tumor cells. Mater Chem Phys., 2011; 128(1–2):83–89.
- 11. Ramgir N, Datta N, Kaur M, Kailasaganapathi S, Debnath AK, Aswal DK, Gupta SK, Metal oxide nanowires for chemiresistive gas sensors: issues, challenges and prospects. Colloids Surf A PhysicochemEng Asp. 2013; 439: 101-116.
- 12. Jani AMM, Losic D, VoelckerNH, Nanoporousanodicaluminium oxide: advances in surface engineering and emerging applications. Prog Mater Sci, 2013; 58:636–704.
- Shalana AE, Rashada MM, Yu Y, Cantub ML, Abdel-Mottaleb MSA, Controlling the microstructure and properties of titaniananopowders for high efficiency dye sensitized solar cells. ElectrochimActa, 2013; 89:469–478.
- 14. Montferrand CD, Huc L, Milosevic I, Russier V, Bonnin D, Motte L, Brioude A, Lalatonne Y, Iron oxide nanoparticles with sizes, shapes and compositions resulting in differentmagnetization signatures as potential labels for multiparametric detection. ActaBiomater, 2013; 9: 6150–6157.
- 15. Ahmadi SJ, Outokesh M, Hosseinpour M, Mousavand T, A simple granulation technique for preparing high-porosity nano copper oxide(II) catalyst beads. Particuology, 2011; 9:480–485.
- 16. Nassar NN, Husein MM, Effect of microemulsion variables on copper oxide nanoparticle uptake by AOT microemulsions. J Colloid InterfSci, 2007; 316:442–450.
- Kao MJ, Lo CH, Tsung TT, Wu YY, Jwo CS, Lin HM, Copperoxide brake nanofluid manufactured using arc-submerged nanoparticle synthesis system. J Alloy Compd, 2007; 434– 435:672–674.
- Chiang CY, Aroh K, Ehrman SH, Copper oxide nanoparticle made by flame spray pyrolysis for photoelectrochemical water splitting e Part I. CuO nanoparticle preparation. Int J Hydrogen Energy, 2012; 37:4871–4879.
- 19. Vijayakumar R, ElgamielR, Diamant Y, Gedanken A, Sonochemical preparation and characterization of nanocrystalline copper oxide embedded in poly (vinyl alcohol) and its effecton crystal growth of copper oxide. Langmuir, 2001; 17:1406–1410.
- 20. Zhang Y, Wang S, Li X, Chen L, Qian Y, Zhang Z, CuOshuttle-like nanocrystals synthesized by oriented attachment. J Cryst Growth, 2006; 291:196–201.

- 21. Wang J, Yang J, Sun J, BaoY,Synthesis of copper oxide nanomaterials and the growth mechanism of copper oxide nanorods. Mater Des, 2004; 25:625–629.
- 22. Iravani S. Green synthesis of metal nanoparticles using plants. Green Chemistry. 2011; 13(10):2638–2650.
- 23. Huang J, Li Q, Sun D, et al. Biosynthesis of silver and gold nanoparticles by novel sundried Cinnamomumcamphora leaf. Nanotechnology, 2007; 18(10):105-104.
- 24. N. Cioffi, N. Ditaranto, L. Torsi, R.A. Picca, E. De Giglio, L. Sabbatini, L. Novello, G. Tantillo, T. Bleve-Zacheo, P.G. Zambonin, Synthesis, analytical characterization and bioactivity of Ag and Cu nanoparticles embedded in poly-vinyl-methyl-ketone films, Anal. Bioanal. Chem. 2005; 382: 1912-1918.
- 25. Xu Q., Y. Zhao, J.Z. Xu, J.J. Zhu, Preparation of functionalized copper nanoparticles and fabrication of a glucose sensor. Sens. Actuators, B Chem. 2006; 114: 379–386.
- 26. Athanassiou E.K., R. N. Grass, W. J. Stark, Large-scale production of carbon-coated copper nanoparticles for sensor applications. Nanotechnology 2006; 17: 1668-1673.
- 27. Pecharromán C., A. Esteban-Cubillo, I. Montero, J. S. Moya, E. Aguilar, J. Santarén, and A. Alvarez, "Monodisperse and corrosión-resistant metallic nanoparticles embedded into sepiolite particles for optical and magnetic applications," J. Am. Ceram. Soc. 2006; 89(10): 3043–3049.
- 28. Honary S, Barabadi H, Fathabad EG, Naghibi F, Green synthesis of copper oxide nanoparticles using penicilliumaurantiogriseum, penicilliumcitrinum and penicilliumwakasmanii. Digest J NanomaterBiostruct, 2012; 7:999–1005.
- 29. Rahman A, Ismail A, JumbiantiD, Magdalena S, Sudrajat H, Synthesis of copper oxide nanoparticles by using Phormidiumcyanobacterium. Indo J Chem2009; 9:355–360.
- 30. Gunalan S, Sivaraj R, Venckatesh R, Aloe barbadensis Miller mediated green synthesis of monodisperse copper oxide nanoparticles: optical properties. SpectrochimActaa 2012; 97:1140–1144.
- 31. Renu Sankar, PerumalManikandan, ViswanathanMalarvizhi, TajudeennasrinFathima, Kanchi Subramanian Shivashangari, VilwanathanRavikumar, Green synthesis of colloidal copper oxide nanoparticles using Carica papaya and its application in photocatalytic dye degradation, Molecular and Biomolecular Spectroscopy, 2014; 121: 746–750.
- 32. Awwad, A.M.1, Albiss, B.A2, Salem N.M., Antibacterial Activity of synthesized Copper Oxide Nanoparticles using MalvasylvestrisLeaf Extract, SMU medical journal, 2015; 2: 91-101.
- 33. VinodVelloraThekkaePadilMiroslavČerník, Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application, International Journal of Nanomedicine, 2013; 8: 889–898.

- 34. Mahmoud Nasrollahzadeh, S. Mohammad Sajadi, Akbar Rostami-Vartooni and MojtabaBagherzadeh, Green synthesis of Pd/CuO nanoparticles by Theobroma cacao L. seeds extract and their catalytic performance for the reduction of 4-nitrophenol and phosphine-free Heck coupling reaction under aerobic conditions, J. Colloid Interface Sci., 2015; 448 (15): 106-113.
- 35. 35. Abboud Y., T. Saffaj, A. Chagraoui, A. El Bouari, K. Brouzi, O. Tanane, B. Ihssane, Biosynthesis, characterization and antimicrobial activity of copper oxide nanoparticles (CONPs) produced using brown alga extract (Bifurcariabifurcata), ApplNanosci, 2014; 4:571–576.
- 36. Amrut. S. Lanje, Satish J. Sharma, Ramchandara B. Pode, Raghumani S. Ningthoujam, Synthesis and optical characterization of copper oxide nanoparticles, Advances in Applied Science Research, 2010; 1 (2): 36-40.
- 37. Raja H, Naika, K. Lingaraju, K. Manjunath, Danith Kumar, G. Nagaraju, D. Suresh, H. Nagabhushana, Green synthesis of CuO nanoparticles using Gloriosasuperba L. extract and their antibacterial activity, Journal of Taibah University for Science, 2015; 9: 7–12.
- Lily Riya and Mary George, Green synthesis of Cuprous Oxide nanoparticles, IJARSE, 2015; 4: 315-322.
- Maryam Bordbar1, Zeinab Sharifi-Zarchi1, Bahar Khodadadi1, Green synthesis of copper oxide nanoparticles/clinoptilolite using Rheum palmatum L. root extract: high catalytic activity for reduction of 4-nitro phenol, rhodamine B, and methylene blue, J Sol-Gel SciTechnol, 2017; 81 (3):724–733.
- 40. Ill-Min Chung , Abdul AbdulRahuman, SampathMarimuthu , Arivarasan Vishnu Kirthi , KarunanithiAnbarasanandGovindasamyRajakumar, Green synthesis of copper nanoparticles using Eclipta prostrate leaves extract and their antioxidant and cytotoxic activities, Experimental and therapeutic medicine, 2017; 14: 18-24.
- 41. Long-Bao Shi, Pei-Fu Tang, Wei Zhang, Yan-Peng Zhao, Li-Cheng Zhang and Hao Zhang, Green synthesis of CuO nanoparticles using Cassia auriculata leaf extract and in vitro evaluation of their biocompatibility with rheumatoid arthritis macrophages (RAW 264.7), Tropical Journal of Pharmaceutical Research January 2017; 16 (1): 185-192.
- 42. Mangesh S. Jadhav,a Sameer. Kulkarni,a Prasad. Raikar,bDelicia A. Barretto,cShyam Kumar. Vootlac and U. S. Raikar,Green Biosynthesis of CuO& Ag-CuO nanoparticles from Malus Domestica leaf extract and evaluation of antibacterial, antioxidant, DNA cleavage activities, New J. Chem., 2018; 42: 204-213.