

Research article

Available online www.ijsrr.org

International Journal of Scientific Research and Reviews

Pesticide Residue Analysis in Mango (*mangifera indica*) Samples From Chittoor District, Andhra Pradesh, India

Sireesha G.¹, Kiranmayi P.¹, Tirupati M. K.² and Karumanchi K. K.²

 ¹Department of Biochemistry, Acharya Nagarjuana university, Nagarjuna Nagar - 522 510, India. E-mail: <u>sirisha bc@yahoo.com; kiranmayikodali@rediffmail.com</u>
² Institute of Frontier technology, RARS, Agricultural University, Tirupati. 517502, India. E-mail: <u>murali_tirupati@rediffmail.com; kirankaarumanchi@gmail.com</u>

ABSTRACT

The present study was aimed to identify fifteen commonly used pesticides in commercial mango fruit samples. The analytical screening was performed by GC-ECD/FPD. The pesticide residues in fruit samples were determined and confirmed by their retention time. The retention time of pesticides was compared with that of the reference standard according to standard guidelines. The data showed that among fifteen mango samples tested, five samples were found to contain the following pesticides: 0.024 μ g g⁻¹ chlorpyrifos (MRL~0.05 μ g g⁻¹), 0.007 μ g g⁻¹ profenofos (MRL~0.05 μ g g⁻¹), 0.013 μ g g⁻¹ hexaconazole (MRL~0.5 μ g g⁻¹), 0.0012 μ g g⁻¹ dimethoate (MRL~1 μ g g⁻¹) and 0.589 μ g g⁻¹ acephate (MRL~2 μ g g⁻¹), respectively. The residue levels were found to be within the permissible limits of these pesticides in fruit samples. The results are part of preliminary screening of the presence of pesticide levels in mango samples and the study shows that the levels are within the maximum permissible limits.

KEYWORDS: Gas Chromatography ECD/FPD, Mangoes, Pesticides, QuEChERS method

*Corresponding author

Kiranmayi P,

Assistant Professor,

Department of Biochemistry, Acharya Nagarjuna University,

Nagarjuna Nagar-522510, India. Phone No: +91 9441748123

E - mail ID: kiranmayikodali@rediffmail.com

INTRODUCTION

Mango (Mangifera indica) belongs to the family of Anacardiaceae and is one of the most popular consuming delicious seasonal fruits in the world and it is known as 'King of Fruits because of its high nutritive values (vitamins, minerals, antiaging factors, antioxidant and anti cancer property)¹. If we look at Indian scenario, Andhra Pradesh stands first in mango production in India^{2,3,4} and also highest pesticide consuming state⁵ because of modern and intensive agricultural practices. Though pesticides are used to enhance the agricultural productivity of crops by protecting from the pests however they are also affecting non-target organisms, thereby causing deleterious effects to the ecosystem and creating a tremendous loss to the biodiversity and human beings⁶. Majority of pesticides (organic /inorganic toxic substances) are seeping into the environment and contaminating our necessary basic needs like air, water, food etc.,^{7, 8} causing neuro-degenerative diseases to human beings^{9, 10} and other possible health effects including respiratory problems such as asthma¹¹, chronic obstructive pulmonary disease (COPD) and lung cancer^{12, 13, 14, 15}, hypersensitivity and also linked with fetal development¹⁶. Although, there are several reports available on pesticide residues in food samples, however for the first time we focused study on the largest mango producing region in Andhra Pradesh, India. In view of the above, the major objective of the present study was to evaluate pesticide residues in mango (Totapuri) sample obtained from different regions of Chittoor district, Andhra Pradesh, India.

MATERIALS AND METHODS

Pesticide Standards and Stock solutions

Analytical reference standards (Dichlorvos 98.8%, Dicofol 99.3%, phosphamidon 94.8%, Hexaconazole 94.8%, Acephate 98.8%, Dimethoate 99.7%, Monocrotophos 99.9%, Deltametrin 95.2%, Malathion 99.1%, Chlorpyrifos 99.7%, Profenofos 95.0%, Cypermethrin 97.8%, and Phosalone 95.2%, Propiconazole 99.1%, Tebuconazole 99.3%) were purchased from Sigma Aldrich (USA). Stock solutions (500 ppm) were prepared by dissolving in N-Hexane.

Sample collection

Fresh mangoes (Tothapuri) from 15 different adjoining villages of Chittoor District, Andhra Pradesh, India were collected. Mango Samples were brought to the laboratory and analyzed.

Sample Preparations

The collected samples were homogenized by a blended homogenizer. The samples were then extracted as per the QuEChERS protocol as reported by Anastassiades *et al.*¹⁷. Briefly, an amount of

15 g of homogenized samples mixed with 30 ml of acetonitrile followed by sonication for 3 min. To this 3 g of activated NaCl was added and shaken gently. The extract was centrifuged for 3 min at 3000 rpm. A 16 ml of the upper layer was transferred into a 50 ml centrifuge tubes which contained 9 g anhydrous Na₂SO₄, shaked for 1 min and kept for sedimentation. The 9 ml upper layer was allowed for clean up with 0.4 g PSA and 1.2 g of MgSO₄ into 15 ml centrifuge tube. The resultant extract was shaken for 1 min and centrifuged at 3000 rpm. The supernatant (2 ml) was collected and evaporated to dryness. The residue was reconstituted with 1 ml of hexane for the analysis. An aliquot of 1 µl extract was injected into GC with ECD and FPD.

Residue analysis

Residue analysis was performed on Shimadzu GC-2010 equipped with ECD and FPD detectors. The capillary column (EB-5) with a stationary phase of 5% phenyl and 95% dimethylpolysiloxane (0.25 mm film thickness). The injector port temperature and detector temperatures were set at 250°C and 280°C, respectively. Nitrogen was selected as a carrier gas with a flow rate of 30 ml min⁻¹, hydrogen was used as a makeup gas with a flow rate of 30 ml min⁻¹ and zero air 60 ml min⁻¹. The oven temperature program was set initially at 240°C with a hold of 3 min, then increase up to 280 at 5°C min⁻¹, hold for 3 min, then increased up to 320 °C at 5 °C min⁻¹ with a hold time of 14 min. The split ratio was set at 1:10, the total run time was 40 min.

Method validation

The validation of the analytical method was performed with respect to the construction of linearity, limit of detection (LOD) and limit of quantification (LOQ). Linearity was established by constructing a calibration graph at five different concentrations in the range of 0.1-1 ppm for all pesticides. The limit of detection (LOD) and limit of quantitation (LOQ) were calculated statistically from the calibration curve and linear regression analysis.

RESULTS AND DISCUSSION

Mango samples from 15 different locations were analyzed for the presence of pesticides residue by GC-ECD/FPD. The peak identification was done by comparing the retention time of standard pesticides with that of samples peaks^{17,18} that illustrates acephate (RT-10.5 min), chlorpyrifos (RT-17.3 min), dimethoate (RT-14.4 min) hexaconazole (RT- 6.1 min) profenofos (RT-20.4 min) with retention times. The calibration graph was constructed for all the pesticides in the range of 0.1-1 ppm and was found to be linear with an acceptable regression coefficient in the range

of 0.980-0.995. The R^2 values for acephate hexaconazole, chlorpyrifos, profenofos, dimethoate was 0.985, 0.985, 0.995, 0.989, and 0.995, respectively as shown in Fig.1.





Furthermore, the results of validation parameters LOD (0.04 mg Γ^1 , 0.03 mg Γ^1 , 0.04 mg Γ^1 , 0.08 mg Γ^1 , 0.05 mg Γ^1) and LOQ (0.12 mg Γ^1 , 0.11 mg Γ^1 , 0.12 mg Γ^1 , 0.26 mg Γ^1 , 0.15 mg Γ^1) for acephate, chlorpyrifos, dimethoate, hexaconazole, profenofos were depicted in Table 1.

Name of the pesticide	Retention Time(min)	LO(mg l ⁻¹)	LOQ(mg l ⁻¹)	Concentratin of Residues (µg g ⁻¹)	MRL (µg g ⁻¹)
Acephate (98.8%)	10.5	0.04	0.12	0.589	2
Chlorpyrifos (99.7%)	17.3	0.03	0.11	0.024	0.05
Dimethoate (99.7%)	14.4	0.04	0.12	0.0012	1
Hexaconazole (94.8%)	6.1	0.08	0.26	0.013	0.5
Profenofos (95.0%)	20.4	0.05	0.15	0.007	0.05

Table 1: Retention time of pesticides along with LOD, LOQ and their concentrations insample against MRLs

Out of fifteen analyzed mango samples, only five samples were contaminated with pesticides. The pesticides detected in the mango samples were as follows: Hexaconazole, Acephate, Chlorpyrifos in sample 1, Acephate, Chlorpyrifos, Profenofos in sample 2, Dimethoate, Acephate, Chlorpyrifos in Sample 5 and Chlorpyrifos in samples 3 and 4 (Fig. 2). The other samples analyzed were not found to contain any pesticides at the analyzed detection limits. The variability in identification levels of pesticide residues in analyzed samples may be attributed to the usage of pesticides and different climatic/growing conditions¹⁹.



Figure 2. GC ECD/ FPD Chromatograms for mango samples showing their retention time

Despite the number of studies available for monitoring the levels of pesticide residue, a study was undertaken to assess the residues of commonly used pesticides, which provides the baseline information for future policy in chemical usage of sample collected. Published reports on pesticide residue analysis in fruits (ber, grapes, and guava) have shown that the residues of the samples were found below its respected MRLs²⁰. In similar lines in the present findings, out of 15 pesticides, only 5 pesticides were identified within the permissible limits and one concentration value of analysed pesticides samples (1-5) against its MRLs ($\sim 2 \mu g g^{-1}$, $\sim 0.05 \mu g g^{-1}$, $\sim 1 \mu g g^{-1}$, $\sim 0.5 \mu g g^{-1} \sim 0.05 \mu g g^{-1}$, respectively) established from Japan Food Chemical Research Foundation 2010²¹ is shown in Table1. This might be due to several factors influencing the fruit such as rain falls²², the time lag

between pesticide spray, harvesting, storage and transportation²³. Secondly, most of the farmers might have enough knowledge about the good agricultural practices. Whereas chlorpyrifos was detected in five samples within the concentration range of 0.017 μ g g⁻¹ – 0.036 ug g⁻¹ (0.034 μ g g⁻¹, $0.024 \ \mu g g^{-1}$, $0.027 \ \mu g g^{-1}$, $0.017 \ \mu g g^{-1}$, $0.036 \ \mu g g^{-1}$), and found below the permissible levels. In this manner, one can assume that there is no apparent risk of the health of consumers, but though they are in permissible limits, previous studies states that continuous exposure to extremely low levels of chlorpyrifos during gestation or early pregnancy may cause detrimental effects on critical periods of development, which has been well documented²⁴. Particularly it was found to be associated with low birth weight²⁵ followed by increased body fat in children²⁶ and finally leads to obesity, cluster of metabolic diseases much later in life^{27, 28}. The results further revealed that the traces of other two pesticides like acephate (0.589 μ g g⁻¹, 0.002 μ g g⁻¹) and profenofos (0.007 μ g g⁻¹) were found, which are considered as unapproved pesticides in fruits according to Food Safety and Standards Authority of India (FSSAI) (2014-15 annual report of the Ministry's Department of Agriculture)²⁹ and people are deliberately bypassing the environmental and health concerns by using these untested pesticides. The fungicide hexaconazole (0.013 μ g g⁻¹) was found below detectable levels. According to Liang *et* $al.^{30}$, hexaconazole exposure might result in thyroid endocrine toxicity and even low concentrations of hexaconazole may affect the development or lead to malformation of Zebrafish larvae. Improper use of hexaconazole may affect the target organism also by reducing the net photosynthesis of plant³¹. Recently, hexaconazole has emerged as a causative agent for oxidative stress in plants³² and ultimately it affects the cellular growth in plants³³. Dimethoate, a widely used organophosphate insecticide, was also detected 0.0012 μ g g⁻¹ in one of the mango samples. There was experimental evidence that other than chlorpyrifos, dimethoate at low levels is also associated with lowered acetylcholinesterase in the brain of rodents³⁴. Few studies have observed that prolonged exposure to low level pesticides can cause harmful health impact on thirteen organ systems³⁵ in human beings. Hence, the present findings indicate that despite a high usage and occurrence of pesticide residues in fruits, it might not be considered as a serious health concern as it was found below permissible limits. However, it is suggested that continuous evaluation of pesticide residues in mango fruits, and further research on changes of metabolic constituents of fruits may throw light on pesticide usage and its impact on fruit quality.

CONCLUSION

Based on the series of samples analyzed in the present study, <50% of the mango samples were contaminated with pesticides and were below permissible limits. These levels of contamination

might vary according to different agricultural practices. However, further experiments need to be conducted to ensure consistent results.

CONFLICT OF INTERESTS

Declared none

ACKNOWLEDGMENTS

We deeply acknowledge the Directorate of Research, Regional agriculture research station, Institute of Frontier Technology for providing the necessary infrastructural facilities to carry out this work. We are thankful to Dr. Mohana Krishna Reddy Mudiam, Principal Scientist, Indian Institute of Chemical Technology, Hyderabad, Telangana for his valuable suggestions. Sireesha G. is thankful to Dr. Ramaiah, Associate Professor, Horticulture Research Station, Anantharaj Pet, YSR District, AP, India. Dr. Narayana, retired scientist, Horticulture Research Station, Anantharaj Pet, YSR District, AP, India for extending their support to carry out this work.

REFERENCES

- Salunkhe DK, Desai BB. Postharvest Biotechnology of Fruits. Business & Economics . Boca Raton, CRC Press: Florida, U.S.A; 1984:1-184.
- Vishwanatha RK, Kumar P. An economic appraisal of mango processing plants of Chittoor District in Andhra Pradesh. Ind J Agri Econ. 2010; 65(2): 277-294.
- Biswas BC, Kumar L. Revolution in mango production: Success stories of some farmers. Fertilizer Marketing News: FAI, New Delhi; March 2011:1-3.
- 4. Shukla PK, Gundappa, AT. Development of sooty moulds in mango orchards in relation to weather parameters and major sucking pests. J Environ Biol. 2017; 38(6): 1293-1300.
- 5. Tulsi B, Sharma JP. Impact of pesticides application in agricultural industry: An Indian Scenario. Int J Agri Food Sci Tech. 2013; 4(8): 817-822.
- Sharma D, Nagpal A, Pakade YB, Katnoria JK. Analytical methods for estimation of organophosphorus pesticide residues in fruits and vegetables. Talanta. 2010; 82(4): 1077-1089.
- Keikothail BM, Spanoghe P, Steurbaut W. Effects of food processing on pesticide residues in fruits and vegetables: A meta-analysis approach. Food Chem Toxicol. 2010; 48(1): 1–6.
- Jolanta F, Tankiewicz M, Biziuk M. Properties and determination of pesticides in fruits and vegetables. Trends Analytical Chem. 2011; 30(6): 814-826.

- Baldi I, Lebailly P, Mohammed Brahim B, Letenneur L, Dartigues JF, Brochard P. Neurodegenerative diseases and exposure to pesticides in the elderly. Am J Epidemiol. 2003; 157(5): 409-14.
- 10. Shweta M, Lahiri DK, Sharma VL, Anand A. Role of early life exposure and environment on neurodegeneration: implications on brain disorders. Transl Neurodegener. 2014; 3(9): 1-14.
- 11. Michael OM. Clinical evaluation of pesticide exposure and poisonings. Lancet. 1997; 349(9059): 1161-1166.
- 12. Margaret DS, Cole D, Abelsohn A, Weir E. Identifying and managing adverse environmental health effects:4. Pesticides. Canadian Med Asso.2002; 166(11): 1431-1436.)
- 13. Alavanja MC, Dosemeci M, Samanic C et al. Pesticides and lung cancer risk in the Agricultural Health Study Cohort. Am J Epidemiol. 2004; 160(9): 876–885.
- 14. Hoppin JA, Valcin M, Henneberger PK et al. Pesticide use and chronic bronchitis among farmers in the agricultural health study. Amer J Ind Med. 2007; 50(12): 969–979.
- 15. Hoppin JA, Umbach DM, London SJ et al. Pesticide use and adult-onset asthma among male farmers in the agricultural health study. Eur Respir. 2009; 34(6): 1296–1303.
- 16. Declerck K, Sylvie R, Christine WV, Katharina M, Camp GV, Schoeters G. Interaction between prenatal pesticide exposure and a common polymorphism in the PON1 gene on DNA methylation in genes associated with cardio- metabolic disease risk- An exploratory study. Clin Epigene. 2017; 9(35): 1-19.
- 17. Anastassiades M, Lehotay SJ, Stajnbaher D, Schenck FJ. Fast and easy multi residue method employing acetonitrile extraction/partitioning and "dispersive solid phase extraction" for the determination of pesticides residues in produce. J AOAC Int. 2003; 86(17): 412–431.
- Lehotay SJ. Quick, easy, cheap, effective, rugged and safe (QuEChERS) approach for determining pesticide residues. In: Vidal J.L.M and Frenich A.G (eds.) Methods in biotechnology. Humana Press: Totowa; 2005:239-261.
- 19. Tomonari Y, Fujita M, Kondo K et al. Factors affecting field-to-field variation in pesticide residue levels in Chinese cabbage and spinach. Pestic Sci. 2013; 38(4): 200-207.
- 20. Kumari B, Madna VK, Kathpal TS. Monitoring of pesticide residues in fruits. Env Monitoring Asses. 2006; 123(1-3): 407-412.
- 21. The Japan Food Chemical Research Foundation, Maximum residue limits (MRLs) list of agricultural chemicals in foods.2010.
- 22. Livia NK, Pasztor Z. Environmental Distribution of acetochlor, atrazine, chlorpyrifos and propisochlor under field conditions. Agric. Food Chem. 2001; 49(8): 3859-3863.

- 23. Holland PT, Hamilton D. Effects of storage and processing on pesticide residues in plant products. Pure Appli Chem. 1994; 66(2): 335-356.
- 24. Timofeeva OAL, Edward D. Lasting Behavioral Consequences of Organophosphate Pesticide Exposure during Develoment: In: Krieger R (eds.). Hayes' Handbook of Pesticide Toxicology. Academic Press: New York; 2010: 837-846.
- 25. Windham G, Fenster L. Environmental contaminants and pregnancy outcomes. Fertil Steril. 2008; 89(2): 111-116.
- Philippe G, Harari R, Barr DB, Debes F. Pesticide Exposure and Stunting as Independent Predictors of Neurobehavioral Deficits in Ecuadorian School Children. Pediatrics. 2005; 117(3): 546-556.
- 27. Meriel W. Human health impacts of exposure to pesticides. Contract ref: 011005, WWF Australia; 2012.
- Nicole EDL, Holloway AC. Early-Life chemical exposures and risk of metabolic syndrome. Diabetes Metab Syndr Obes: Targets and Therapy. 2017; 10: 101-109.
- 29. Ananya T, Priya OS. Report highlights use of non approved pesticides in food items in India. Down To Earth (DTE) News, New Delhi; October 2015.
- Liang YU, Mengli CC, Yihua L., Wenjun G, Guonian Z. Thyroid endocrine disruption in zebrafish larvae following exposure to hexaconazole and tebuconazole. Aqua Toxicol. 2013; 138: 35-42.
- 31. Xia XY, Huang YY, Wang L et al. Pesticides induced depression of photosynthesis was alleviated by 24- epi brassinolide pretreatment in *Cucumis sativus* L. Pestic Biochem Physiol. 2006; 86(1): 42-48.
- 32. Bourouhou M, Boudouna BE, Mokhtar B. The impact of a hexaconazole fungicide on agronomic, biochemical parameters and yield components of green beans *Phaseolus Vulgaris* cv. Djedida. Int J Adv Eng Manag. 2017; 2(6): 146-152.
- 33. Pallavi S, Ambuj BJ, Dubey RSR, Pessarakli M. Reactive Oxygen Species, Oxidative Damage and Antioxidative Defense Mechanism in Plants under Stressful Conditions. J Botany. 2012; 10 (1155): 1-26.
- 34. Srivastava MK, Raizada RB. Development effect of technical dimethoate in rats: maternal and fetal toxicity evaluation. Indian J Exp Biol. 1996; 34(4): 329-333.
- 35. Roni AN, Jennifer CH, Linnea IL, Kathleen D, Anne CR, Keeve EN. A comparative study of allowable pesticide residue levels on produce in the United States. Global Health. 2012; 8(1):

^{2.}